



Advanced NMR: Metabolite ID by NMR

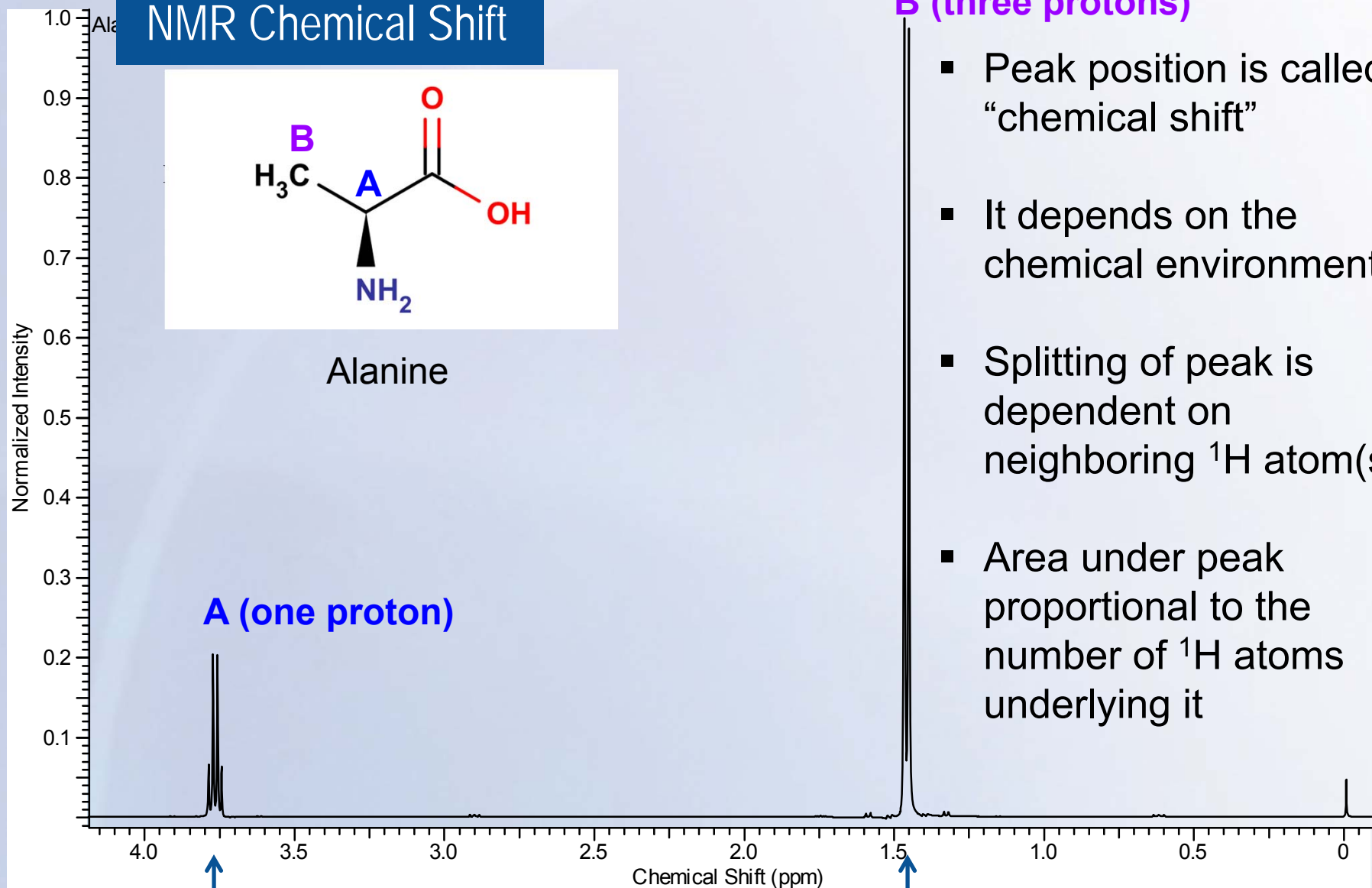
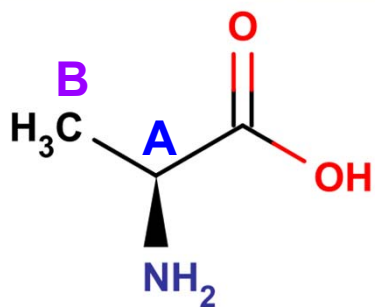
UAB Metabolomics Training Course

June 14-18, 2015

Wimal Pathmasiri, Rodney Snyder
NIH Eastern Regional Comprehensive Metabolomics Resource Core
(RTI RCMRC)

- Information that NMR Spectroscopy data can provide
 - Chemical shift, J-coupling, chemical structure
- Available NMR methods
 - 1D NMR: ^1H , ^{13}C , ^{15}N , ^{31}P
 - 2D NMR: COSY, TOCSY, HSQC, HMBC, NOESY, INADEQUATE
 - Selective 1D: 1D TOCSY, 1D HSQC
- Spectral editing methods
 - CPMG, Diffusion, JRES, DEPT (DEPT 45, DEPT 90, DEPT 135)
- NMR Libraries, software, and databases
 - AMIX, BBREFCODE (Bruker), BATMAN, Chenomx, COLMAR, HMDB, BMRB, Birmingham Metabolite Library, NMR Shift DB
- Other complementary methods
 - Eg. STOCSY, STORM, RANSY
 - MUMA Package (R Based) for STOCSY and STORM
- Tagging, Spike-in of metabolites, Predicting Spectra

NMR Chemical Shift

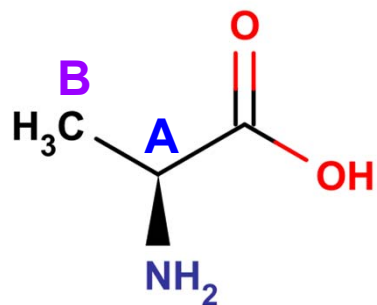


B (three protons)

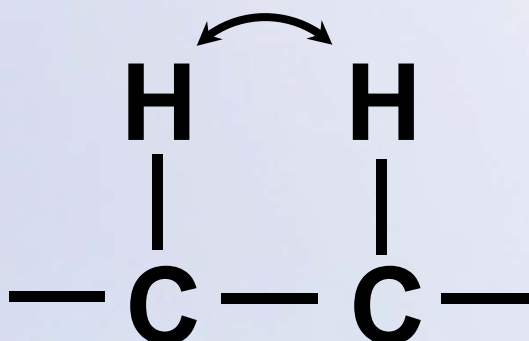
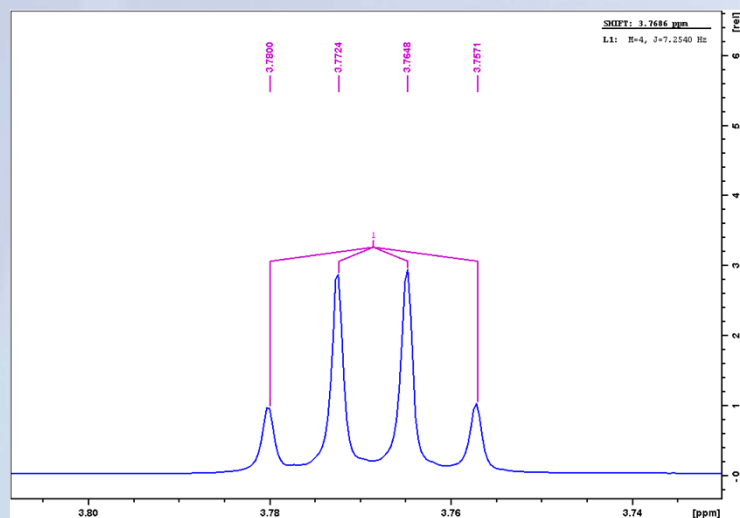
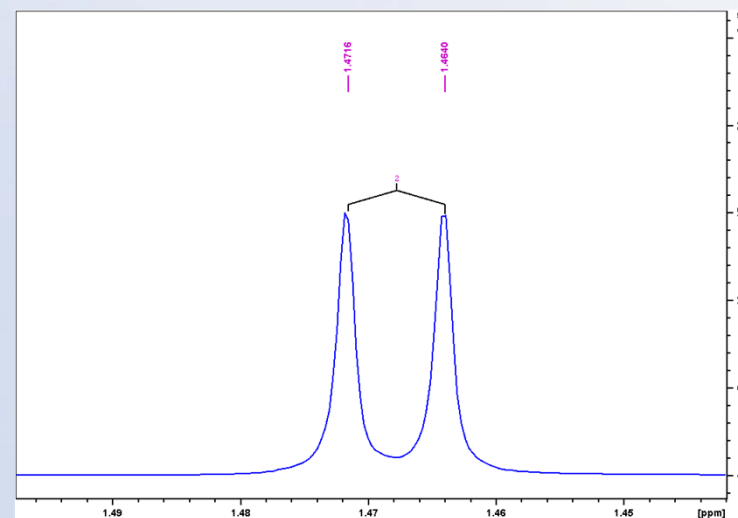
- Peak position is called “chemical shift”
- It depends on the chemical environment
- Splitting of peak is dependent on neighboring ^1H atom(s)
- Area under peak proportional to the number of ^1H atoms underlying it

Chemical shift of CH
4 lines (quadrat)

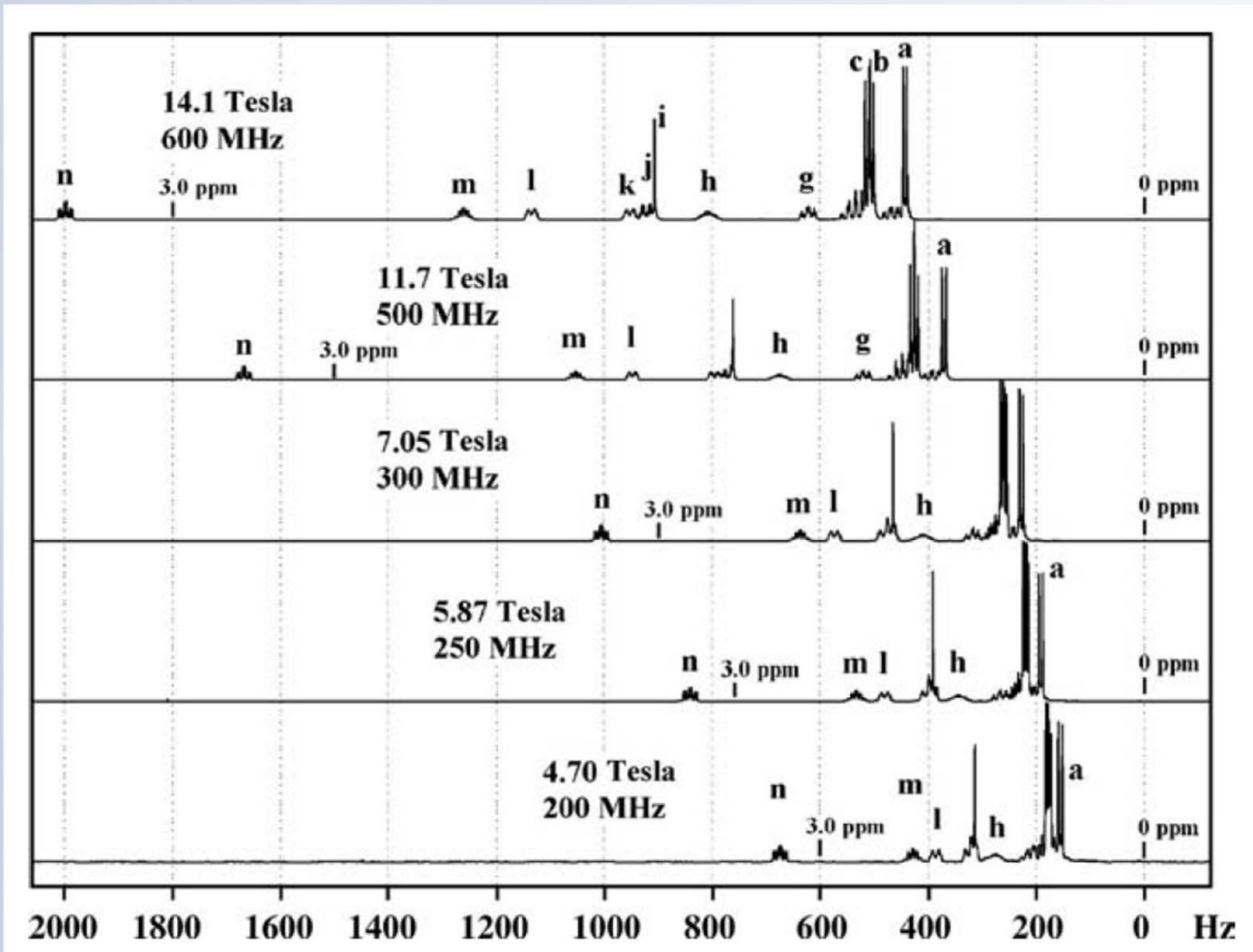
Chemical shift of CH_3
2 lines (doublet)



Alanine

 $J = 7.2 \text{ Hz}$  $J = 7.2 \text{ Hz}$ 

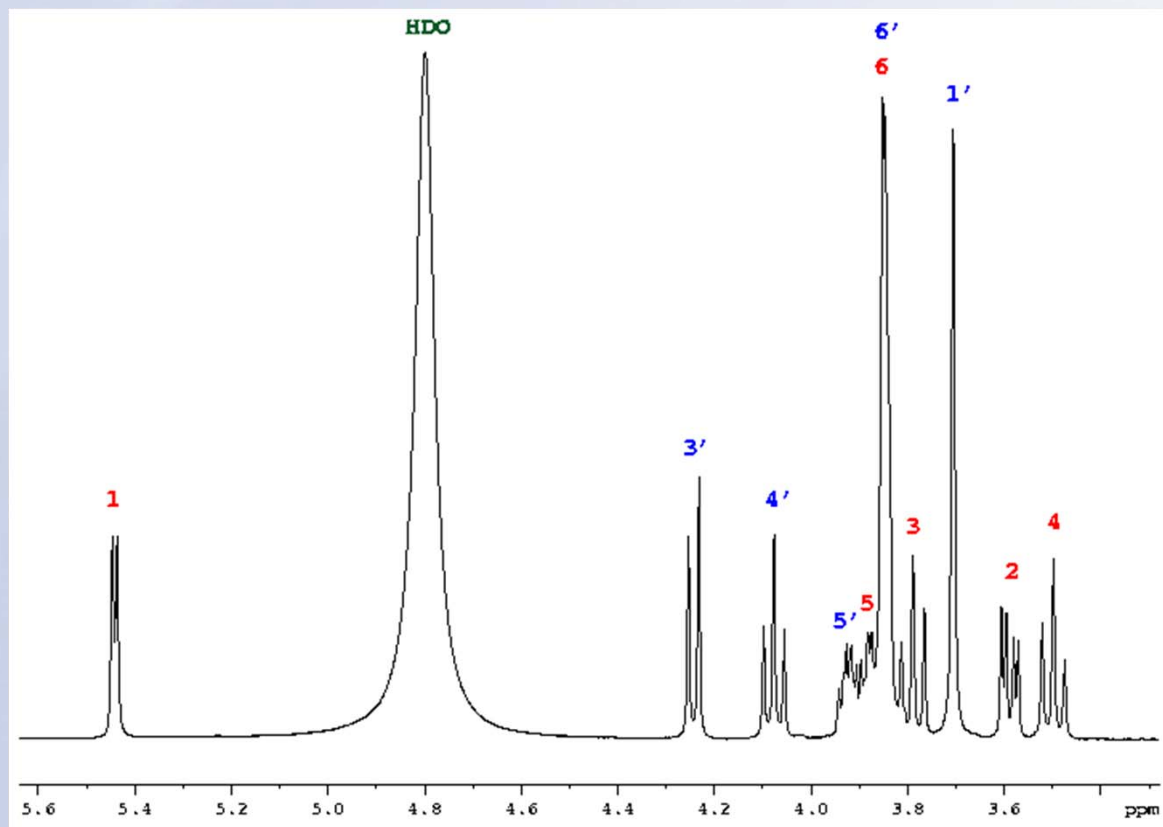
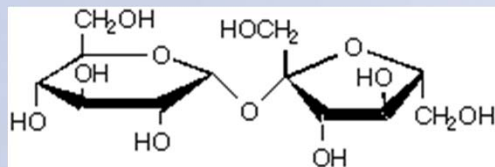
Dispersion of NMR Signal with Magnetic Field Strength



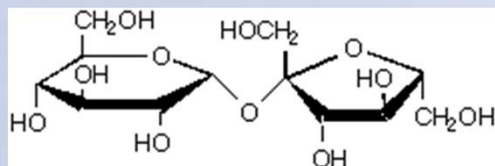


1D and 2D NMR Methods

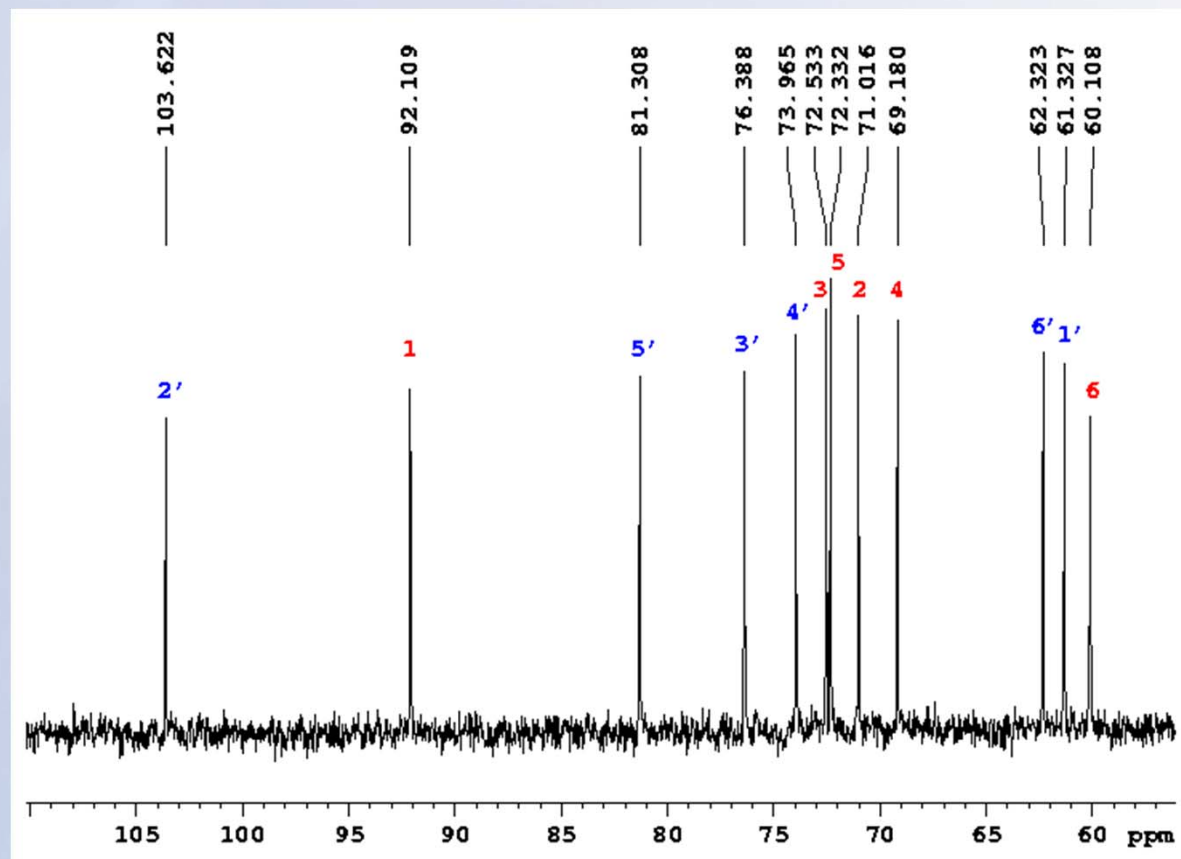
^1H NMR: Sucrose



NMRGuide 4.3 - TOPSPIN 3.0, Bruker BioSpin

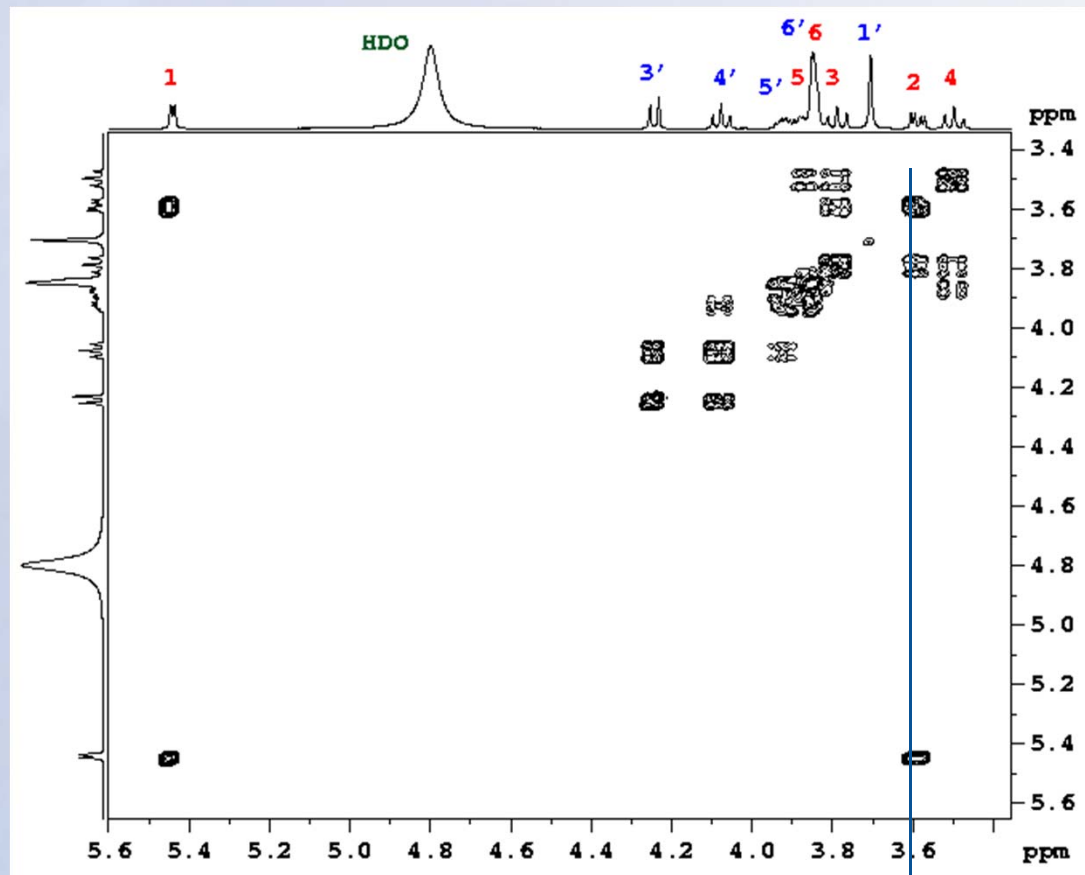
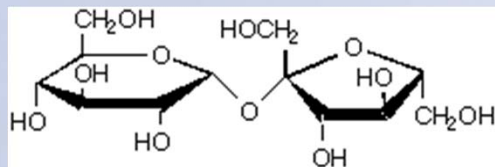


^{13}C NMR: Sucrose



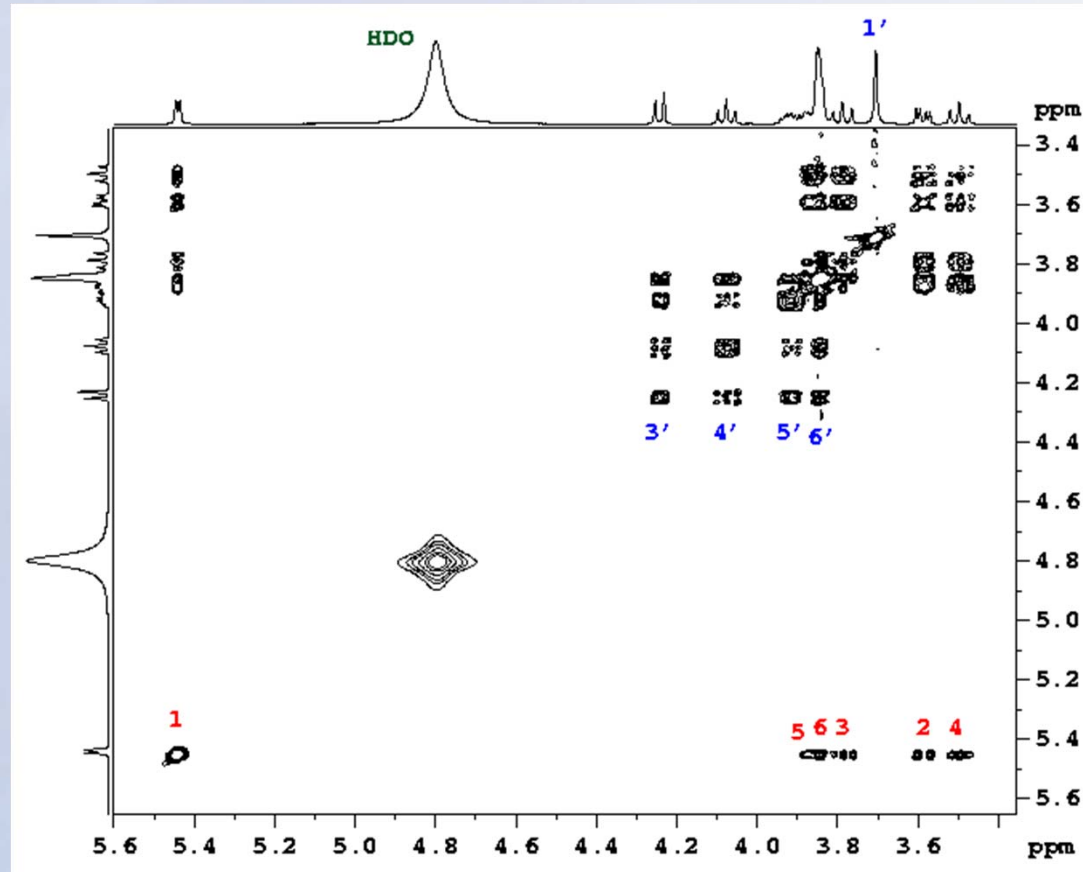
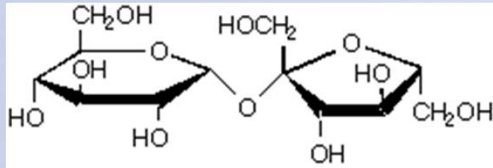
NMRGuide 4.3 - TOPSPIN 3.0, Bruker BioSpin

COSY: Sucrose

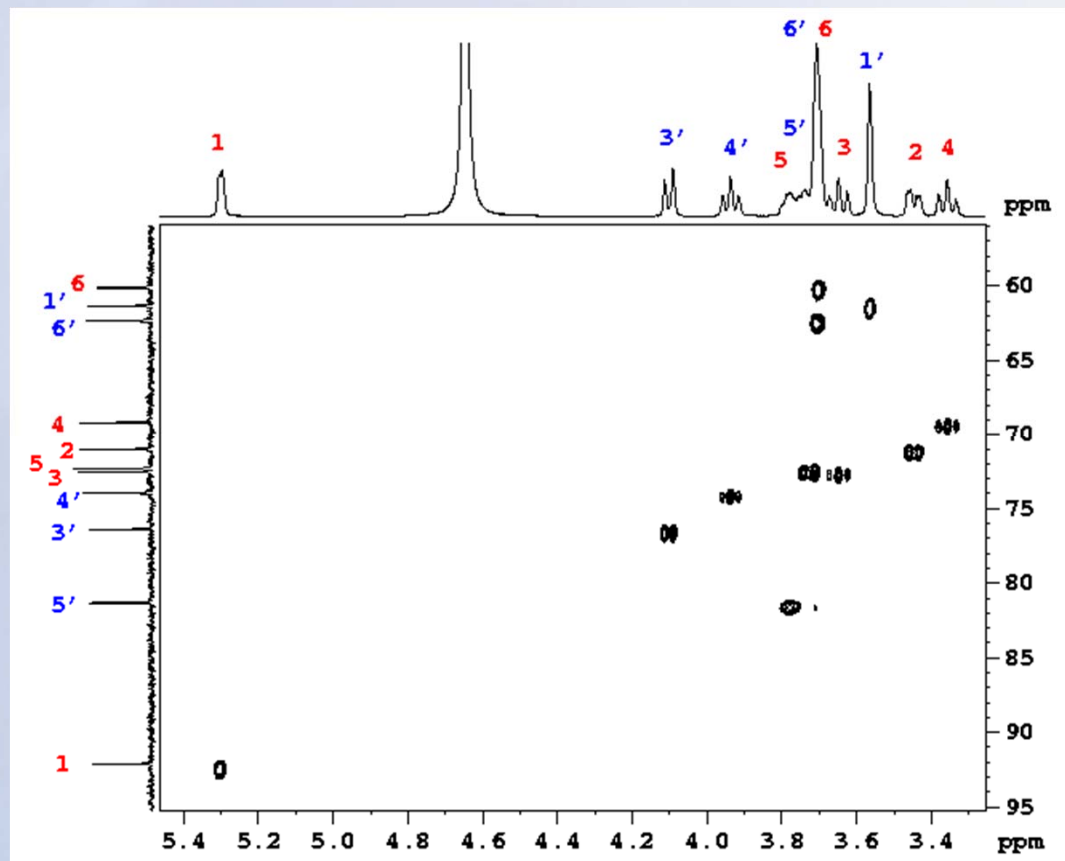
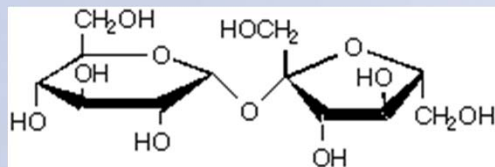


NMRGuide 4.3 - TOPSPIN 3.0, Bruker BioSpin

TOCSY: Sucrose

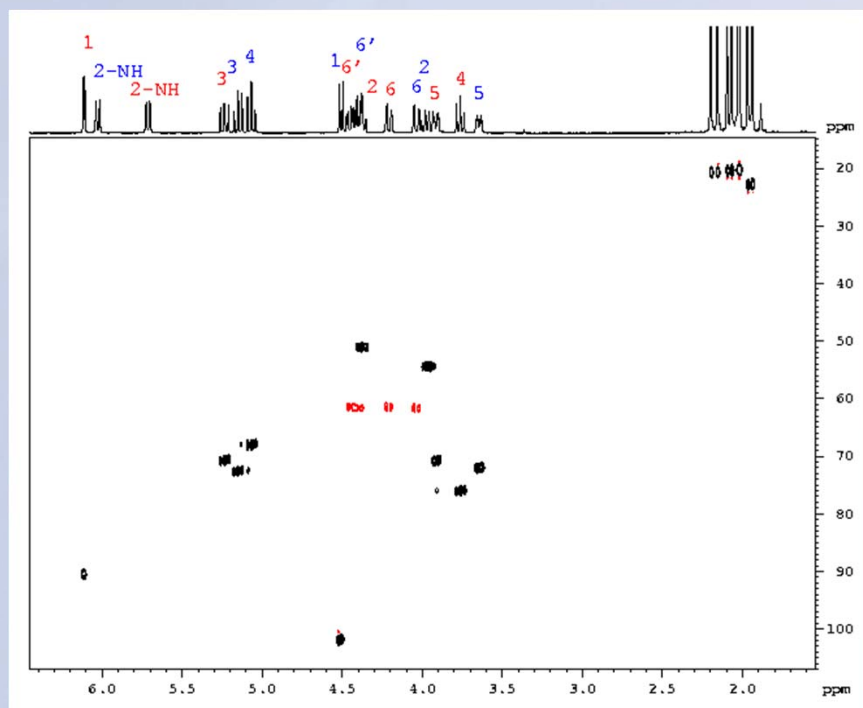
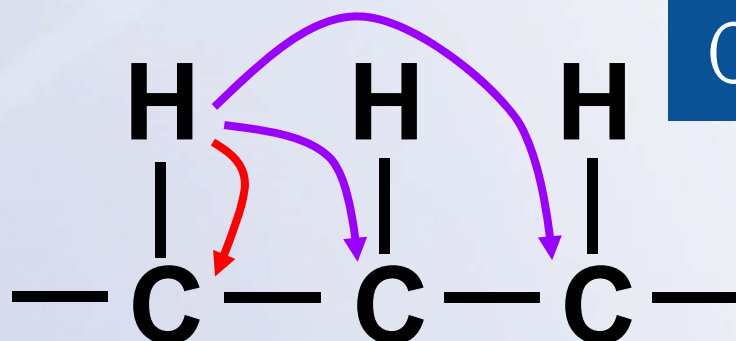
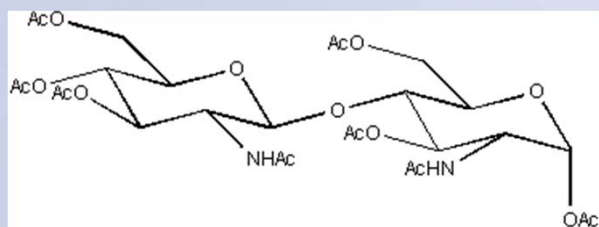


HSQC: Sucrose

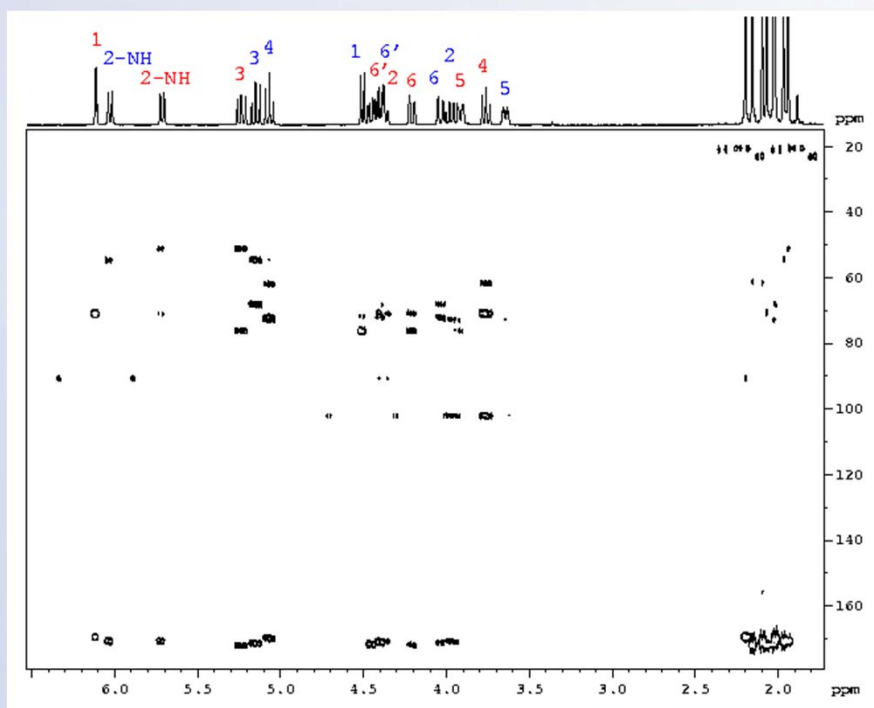


NMRGuide 4.3 - TOPSPIN 3.0, Bruker BioSpin

Chentobiose

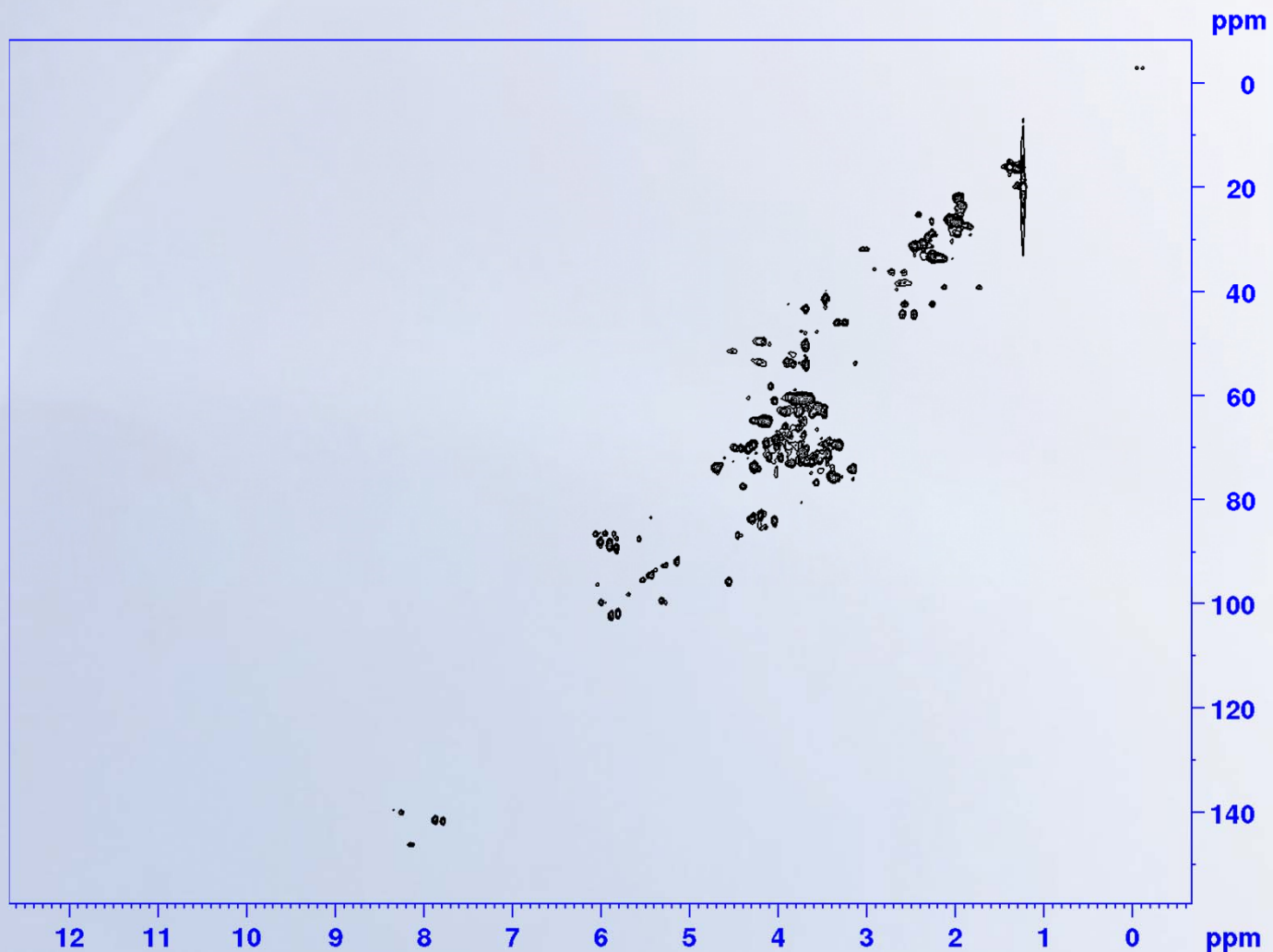


HSQC ($^1J_{CH}$)



HMBC ($^2J_{CH}$, $^3J_{CH}$)

^1H - ^{13}C HSQC Spectrum of Cell Extract

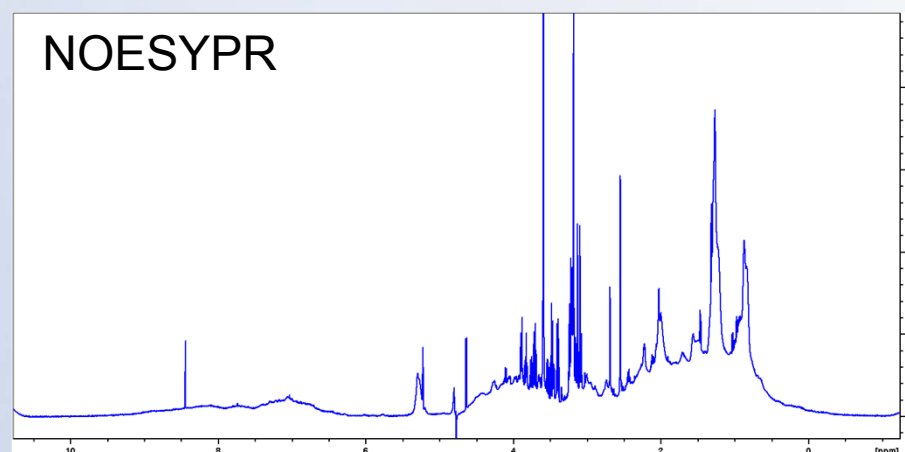
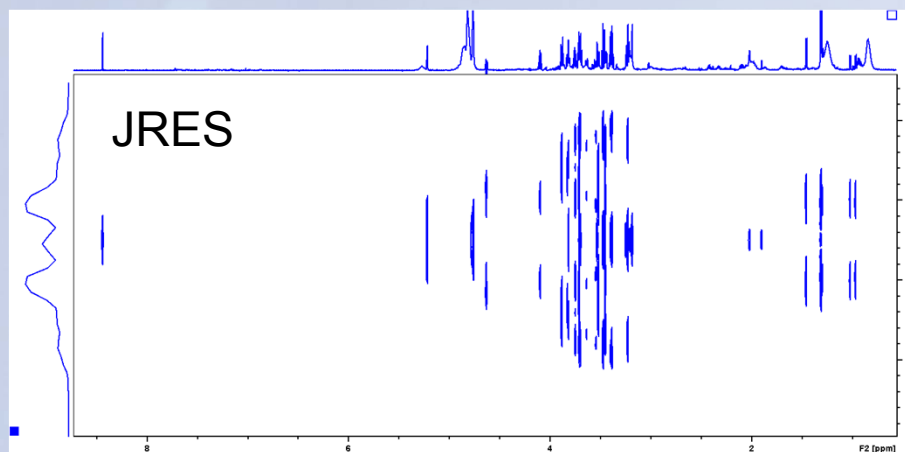
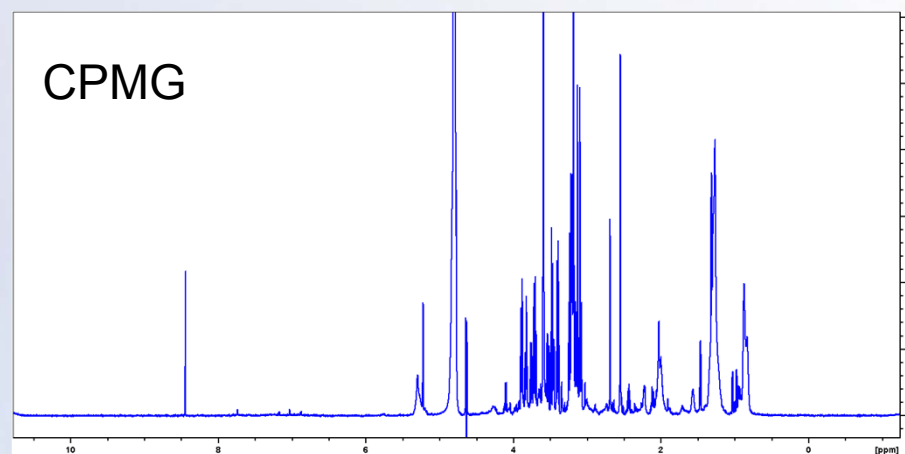




Spectral Editing

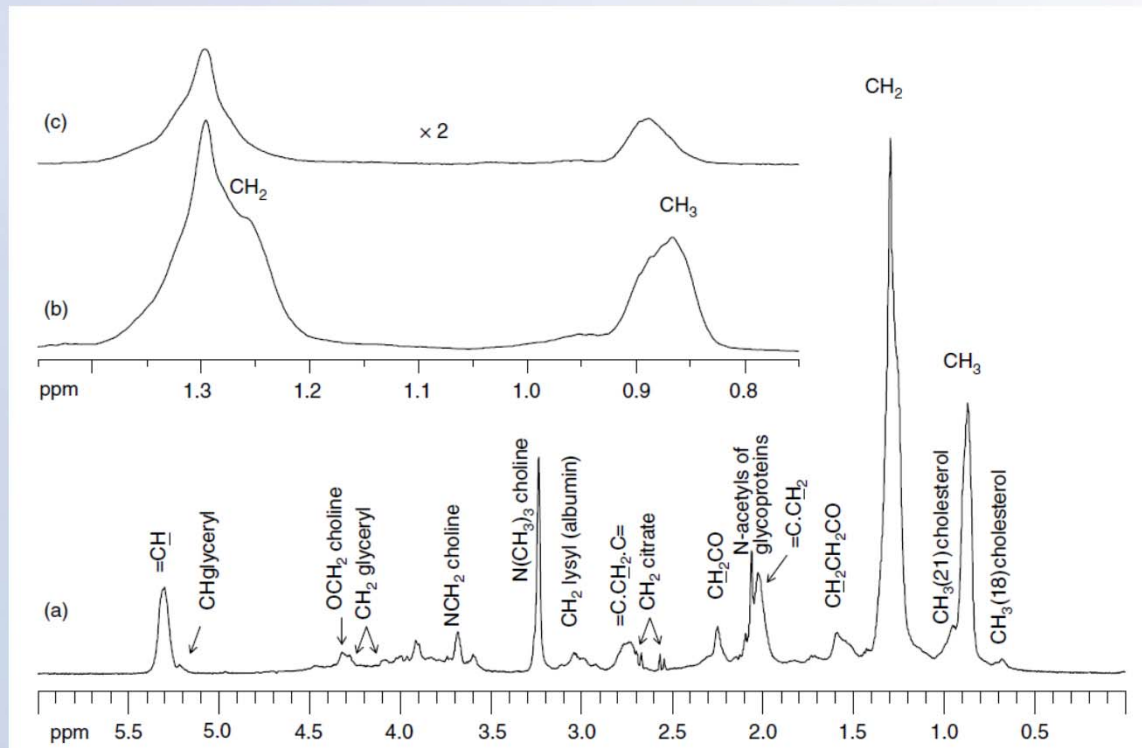
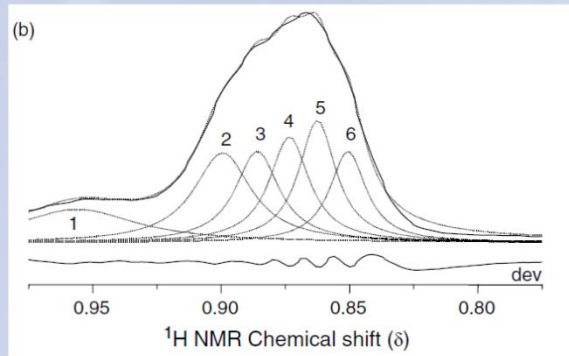
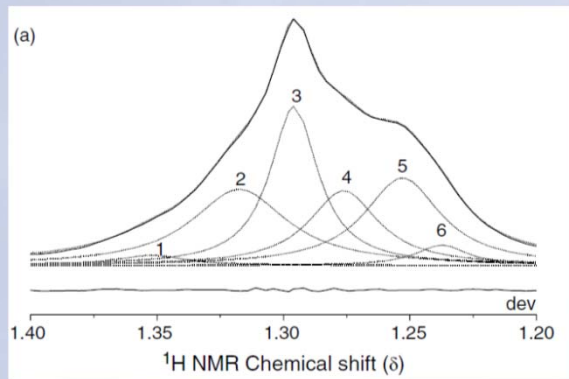
Spectral Editing

- Relaxation editing
 - CPMG Pulse sequence
- J- Modulated
 - J-RES



Analysis of Lipoproteins

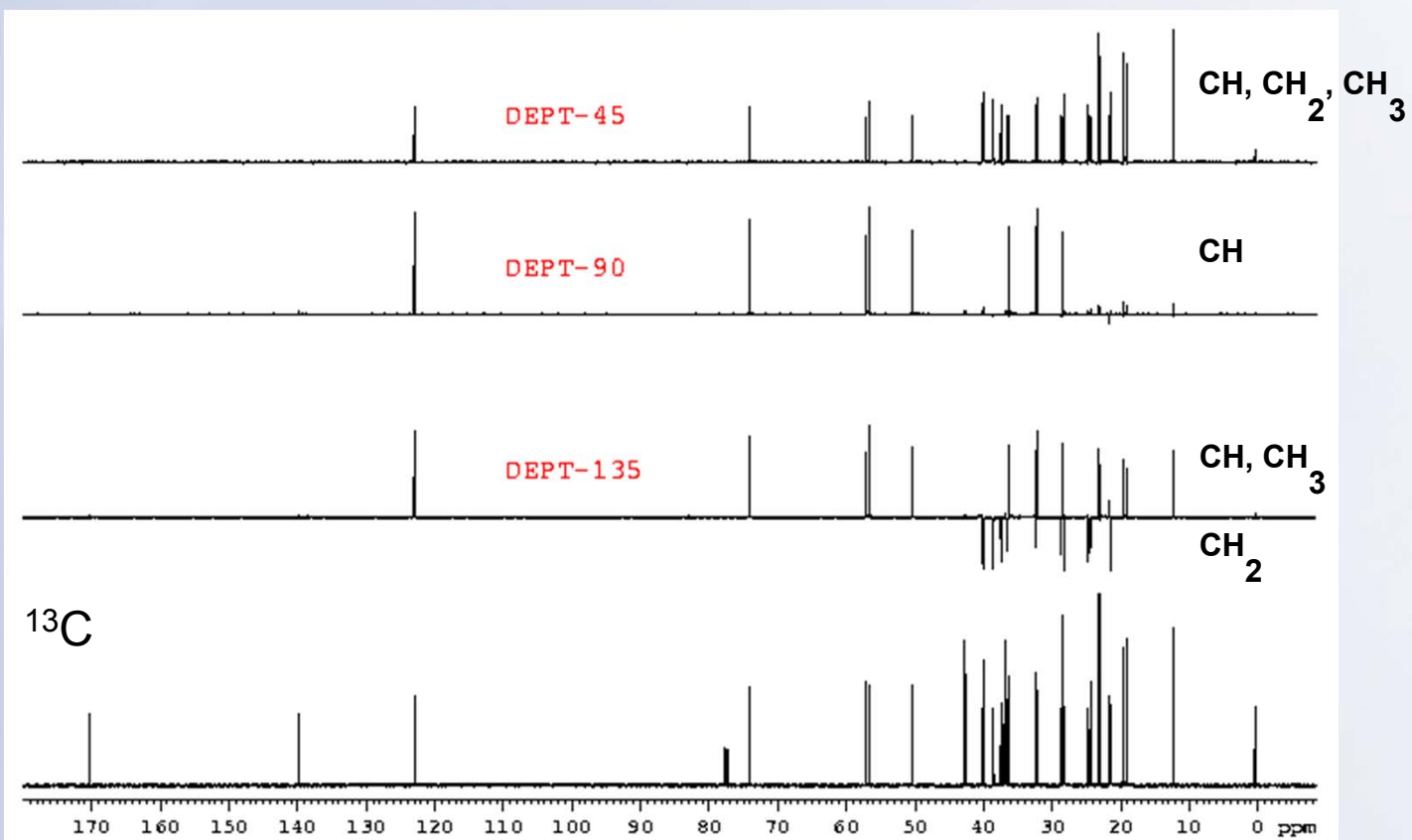
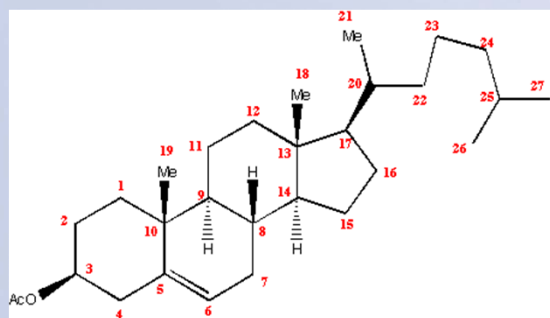
- Lipoproteins are classified based on their size and density (VLDL, LDL, HDL)
- Lipoproteins can be assigned to these sub fractions by deconvoluting the CH_3 and $(\text{CH}_2)_n$ in diffusion edited spectra



Diffusion Edited NMR Spectra

Peak	δ (ppm)	Width (Hz)	D ($\text{cm}^2 \text{s}^{-1} \times 10^7$) ^a	Area (%) ^b	Assignment
$(\text{CH}_2)_n$ 1	1.353	19.7	2.72	2.3	LDL + VLDL
$(\text{CH}_2)_n$ 2	1.317	28.0	1.98	26.4	VLDL
$(\text{CH}_2)_n$ 3	1.296	14.0	1.85	27.9	VLDL
$(\text{CH}_2)_n$ 4	1.276	17.8	3.15	15.5	LDL
$(\text{CH}_2)_n$ 5	1.255	20.5	5.19	19.9	HDL(60.6%) + LDL(39.4%)
$(\text{CH}_2)_n$ 6	1.240	18.4	5.96	7.9	HDL
CH_3 1	0.956	33.9	3.77	15.8	VLDL + HDL
CH_3 2	0.899	16.4	1.70	20.6	VLDL
CH_3 3	0.886	12.4	1.84	15.9	VLDL
CH_3 4	0.873	11.3	3.07	16.7	LDL
CH_3 5	0.862	10.5	4.51	17.9	HDL(40.8%) + LDL(59.2%)
CH_3 6	0.851	10.2	7.11	13.1	HDL

DEPT: Cholesterol





NMR Libraries, Software, and Databases

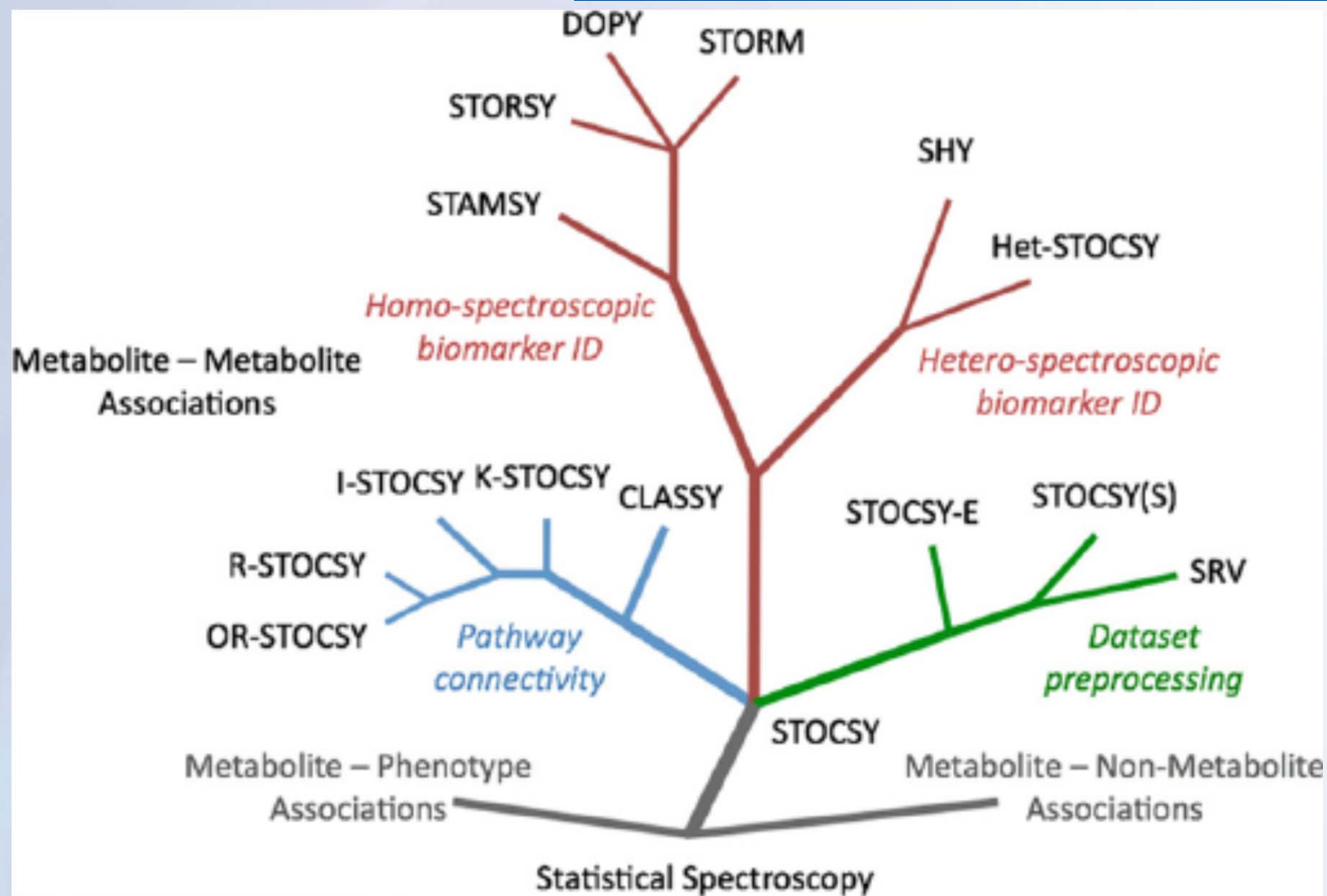
NMR Libraries and Databases

- Continuously emerging
- Databases
 - HMDB (<http://www.hmdb.ca/>)
 - Birmingham Metabolite Library (<http://www.bml-nmr.org/>)
 - BMRB (<http://www.bmrwisc.edu/>)
 - NMRShift DB (<http://nmrshiftdb.nmr.uni-koeln.de/>)
- Online Software
 - COLMAR (<http://spin.ccic.ohio-state.edu/>)
- Standalone Software
 - Chenomx (<http://www.chenomx.com/>)
 - AMIX/ ASSURE/BBREFCODE
<https://www.bruker.com/products/mr/nmr/nmr-software/software/amix/overview.html>
 - BATMAN (<http://batman.r-forge.r-project.org/>)
 - CCPN Metabolomics (<http://www.ccpn.ac.uk/collaborations/metabolomics>)
 - rNMR (link)



Other Complementary methods

Statistical Spectroscopic Tools

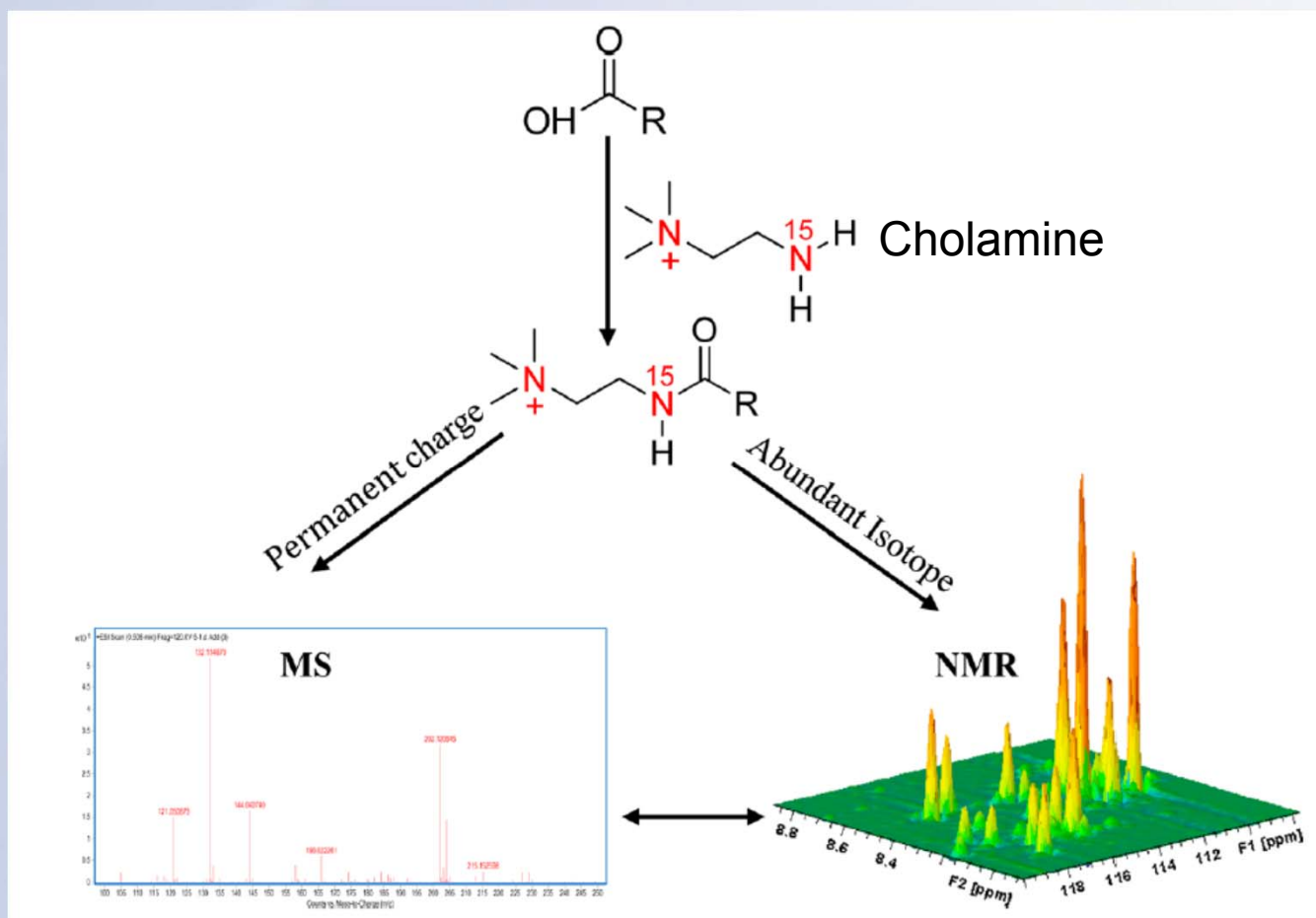


Robinette, S. L., J. C. Lindon and J. K. Nicholson (2013). "Statistical spectroscopic tools for biomarker discovery and systems medicine." *Anal Chem* 85(11): 5297-5303.



Tagging

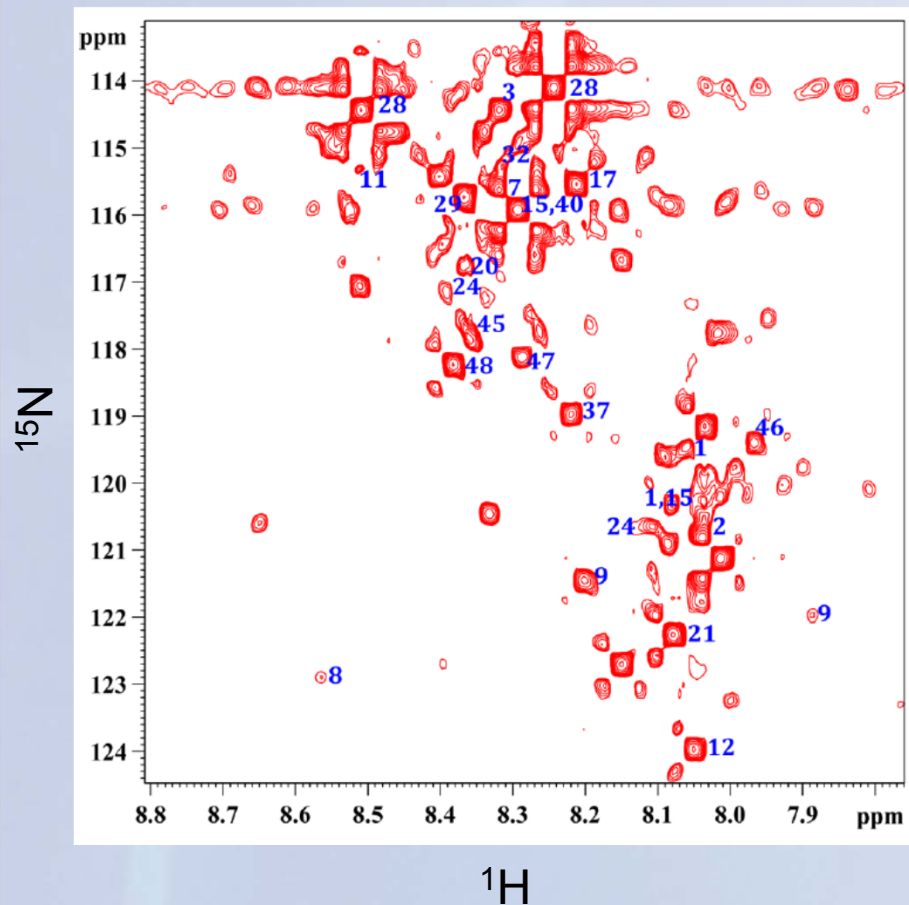
Smart Tagging



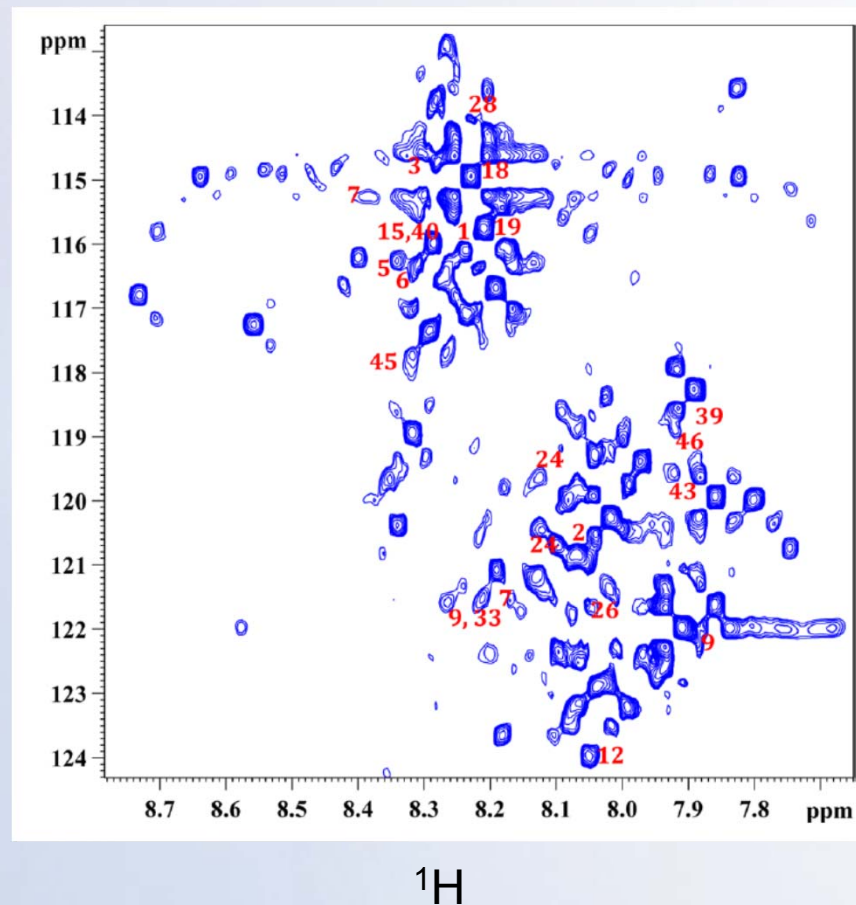
Tayyari, F., G. A. Gowda, H. Gu and D. Raftery (2013). " ^{15}N -cholamine--a smart isotope tag for combining NMR- and MS-based metabolite profiling." *Anal Chem* **85**(18): 8715-8721

^1H - ^{15}N HSQC

Human Serum



Human Urine



Tayyari, F., G. A. Gowda, H. Gu and D. Raftery (2013). " ^{15}N -cholamine--a smart isotope tag for combining NMR- and MS-based metabolite profiling." *Anal Chem* **85**(18): 8715-8721

References

- Beckonert, O., H. C. Keun, T. M. Ebbels, J. Bundy, E. Holmes, J. C. Lindon and J. K. Nicholson (2007). "Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts." *Nat Protoc* 2(11): 2692-2703.
- Cloarec, O., M. E. Dumas, J. Trygg, A. Craig, R. H. Barton, J. C. Lindon, J. K. Nicholson and E. Holmes (2005). "Evaluation of the Orthogonal Projection on Latent Structure Model Limitations Caused by Chemical Shift Variability and Improved Visualization of Biomarker Changes in ^1H NMR Spectroscopic Metabonomic Studies." *Analytical Chemistry* 77(2): 517-526.
- Lindon, J. C. and J. K. Nicholson (2008). "Spectroscopic and Statistical Techniques for Information Recovery in Metabonomics and Metabolomics." *Annual Review of Analytical Chemistry* 1(1): 45-69.
- Posma, J. M., I. Garcia-Perez, M. De Iorio, J. C. Lindon, P. Elliott, E. Holmes, T. M. Ebbels and J. K. Nicholson (2012). "Subset optimization by reference matching (STORM): an optimized statistical approach for recovery of metabolic biomarker structural information from ^1H NMR spectra of biofluids." *Anal Chem* 84(24): 10694-10701.
- Robinette, S. L., J. C. Lindon and J. K. Nicholson (2013). "Statistical spectroscopic tools for biomarker discovery and systems medicine." *Anal Chem* 85(11): 5297-5303.
- Sands, C. J., M. Coen, T. M. Ebbels, E. Holmes, J. C. Lindon and J. K. Nicholson (2011). "Data-driven approach for metabolite relationship recovery in biological ^1H NMR data sets using iterative statistical total correlation spectroscopy." *Anal Chem* 83(6): 2075-2082.
- Wei, S., J. Zhang, L. Liu, T. Ye, G. A. Gowda, F. Tayyari and D. Raftery (2011). "Ratio analysis nuclear magnetic resonance spectroscopy for selective metabolite identification in complex samples." *Anal Chem* 83(20): 7616-7623.
- Tayyari, F., G. A. Gowda, H. Gu and D. Raftery (2013). " ^{15}N -cholamine--a smart isotope tag for combining NMR- and MS-based metabolite profiling." *Anal Chem* 85(18): 8715-8721.



Chenomx Exercise

Chenomx Library

1,3-Dihydroxyacetone, 1,3-Dimethylurate, 1,6-Anhydro- β -D-glucose, 1,7-Dimethylxanthine, 1-Methylnicotinamide, 2'-Deoxyadenosine, 2'-Deoxyguanosine, 2'-Deoxyinosine, 2-Amino adipate, 2-Aminobutyrate, 2-Ethylacrylate, 2-Furoate, 2-Hydroxy-3-methylvalerate, 2-Hydroxybutyrate, 2-Hydroxyglutarate, 2-Hydroxyisobutyrate, 2-Hydroxyisocaproate, 2-Hydroxyisovalerate, 2-Hydroxyphenylacetate, 2-Hydroxyvalerate, 2-Methylglutarate, 2-Octenoate, 2-Oxobutyrate, 2-Oxocaproate, 2-Oxoglutarate, 2-Oxoisocaproate, 2-Oxovalerate, 2-Phosphoglycerate, 3,4-Dihydroxymandelate, 3,5-Dibromotyrosine, 3-Aminoisobutyrate, 3-Chlorotyrosine, 3-Hydroxy-3-methylglutarate, 3-Hydroxybutyrate, 3-Hydroxyisovalerate, 3-Hydroxymandelate, 3-Hydroxyphenylacetate, 3-Indoxylsulfate, 3-Methyl-2-oxovalerate, 3-Methyladipate, 3-Methylxanthine, 3-Phenyllactate, 3-Phenylpropionate, 4-Aminobutyrate, 4-Aminohippurate, 4-Hydroxy-3-methoxymandelate, 4-Hydroxybutyrate, 4-Hydroxybutyrate, 4-Hydroxyphenylacetate, 4-Hydroxyphenyllactate, 4-Pyridoxate, 5,6-Dihydrothymine, 5,6-Dihydrouracil, 5-Aminolevulinatate, 5-Hydroxyindole-3-acetate, 5-Hydroxylysine, 5-Methoxysalicylate, Acetaldehyde, Acetamide, Acetaminophen, Acetate, Acetoacetate, Acetone, Acetylsalicylate, Adenine, Adenosine, Adipate, Alanine, Allantoin, Alloisoleucine, Anserine, Arginine, Argininosuccinate, Asparagine, Aspartate, Benzoate, Betaine, Biotin, Butyrate, Butyrate, Caffeine, Caprate, Caprylate, Carnitine, Carnosine, Choline, Cinnamate, Citrate, Citrulline, Creatine, Creatinine, Cysteine, Cystine, Cytidine, Cytosine, DSS (Chemical Shift Indicator), Dimethylamine, Epicatechin, Ethanol, Ethanolamine, Ethylene glycol, Ethylmalonate, Ferulate, Formate, Fructose, Fucose, Fumarate, Galactarate, Galactitol, Galactonate, Galactose, Gentisate, Glucarate, Glucose, Glutamate, Glutamine, Glutarate, Glutaric acid monomethyl ester, Glutathione, Glycerate, Glycerol, Glycine, Glycolate, Glycylproline, Guanidoacetate, Guanine, Hippurate, Histidine, Homocitrulline, Homocystine, Homogentisate, Homoserine, Homovanillate, Hypoxanthine, Ibuprofen, Imidazole, Indole-3-acetate, Inosine, Isobutyrate, Isocaproate, Isocitrate, Isoleucine, Isopropanol, Isovalerate, Kynurenate, Kynurenine, Lactate, Lactose, Leucine, Levulinatate, Lysine, Malate, Maleate, Malonate, Mannitol, Mannose, Methanol, Methionine, Methylamine, Methylguanidine, Methylmalonate, Methylsuccinate, N,N-Dimethylformamide, N,N-Dimethylglycine, N-Acetylaspartate, N-Acetylglutamate, N-Acetylglutamine, N-Acetyllysine, N-Carbamoyl- β -alanine, N-Carbamoylaspartate, N-Isovaleroylglycine, NAD⁺, Niacinamide, Nicotinate, O-Acetylcarnitine, O-Phosphocholine, O-Phosphoethanolamine, O-Phosphoserine, Ornithine, Oxalacetate, Oxypurinol, Pantothenate, Phenol, Phenylacetate, Phenylacetylglutamine, Phenylalanine, Pimelate, Proline, Propionate, Propylene glycol, Protocatechuate, Pyridoxine, Pyroglutamate, Pyruvate, Quinolinatate, Riboflavin, Ribose, S-Adenosylhomocysteine, S-Sulfocysteine, Salicylate, Salicylurate, Sarcosine, Serine, Suberate, Succinate, Succinylacetone, Sucrose, Tartrate, Taurine, Theophylline, Threonate, Threonine, Thymine, Thymol, Tiglylglycine, Trigonelline, Trimethylamine, Trimethylamine N-oxide, Tryptophan, Tyramine, Tyrosine, Uracil, Urea, Uridine, Urocanate, Valerate, Valine, Valproate, Vanillate, Xanthine, Xanthosine, Xylose, cis-Aconitate, myo-Inositol, o-Cresol, p-Cresol, trans-4-Hydroxy-L-proline, trans-Aconitate, β -Alanine, π -Methylhistidine, τ -Methylhistidine

- Over 320 metabolites

- pH sensitive library of 1H NMR Spectra

- Customizable

Chenomx Exercise

- Save the folder called “Chenomx_Tutorial.zip into your computer
 - Sample files
 - Chenomx NMR Suite Tutorial.pdf

- We will use Processor and Profiler in the exercise

- Processor
 - Sample.fid

- Profiler
 - Basic Start, Basic End
 - Advanced Start, Advanced End
 - Batch fitting

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